# Leptospira, Giardia spp., and Cryptosporidium spp. across a Land Use Gradient

Kalvanii Kennedy\*, OMS-II, Kathleen Shea Hayes\*, Meghan Blackledge\*, Alejandra Orteja\*. Brianna Beechler, DVM, PhD, Rhea Hanselmann, DVM, MPVM, PhD, Michelle Steinauer, PhD 1 Basic Medical Sciences, Western University of Health Sciences COMP-Northwest, Lebanon,

**OR** 

2 Carlson College of Veterinary Medicine, Oregon State University, Corvallis, OR \*Equally contributing authors

# INTRODUCTION

OF HEALTH SCIENCE

College of Osteopathic Medicine of the Pacific **COMP-Northwest** 

Understanding factors that drive the distribution of waterborne pathogens can help mitigate infection risk. This project aimed to determine the distribution of three waterborne pathogens, Leptospira, Giardia, and Cryptosporidium along a land use gradient in two watersheds in Oregon. Understanding how these waterborne pathogens are transmitted is key to understanding distribution. Giardia and Cryptosporidium are both transmitted through feces while Leptospira is transmitted through urine. Escherichia coli is a gram-negative bacillus that is part of the normal GI flora in the lower intestinal tract and can be used as a measurement of fecal contamination in water. This research is an ongoing One Health Initiative Project with data available from last year to review and analyze for any trends.









Figure 1 High-resolution scanning Figure 2 Giardia duodenalis electron micrograph of Leptospira interrogans trophozoites in Giemsa stai

Figure 3 Cryptosporidum parvum electron microscope

### OBJECTIVES

Understand the relationship between waterborne pathogen distribution and the land use patterns

Determine if the presence of waterborne pathogens is correlated with water quality as measured by Total Dissolved Solids (TDS) and E.coli colony counts Determine if the abundance of Escherichia coli in water samples will positively

correlate with the presence of waterborne pathogens.

## STUDY DESIGN

Water was sampled from 24 sites along Marys River in the Willamette Valley, Oregon and 15 sites along White River in north-central Oregon in low, medium, and high disturbance areas

Sites were designated as low, medium, and high disturbance based on surrounding land use

Temperature, total dissolved solids (TDS), and pH was measured at each site with a water meter, and E. coli was isolated and enumerated at each site as a measure of water quality

1 L of water was collected at each site for pathogen detection and filtered with a 2 µm Nalgene filter.

Filters were divided and part was processed so that the filtered debris was preserved in a formalin buffer for an immunofluorescence assay

DNA was extracted from the other portion of the filter for digital droplet PCR (dPCR) detection

Merifluor® sample kits were used to test for the presence or absence of Giardia and Cryptosporidium in the formalin preserved filter debris using direct immunofluorescence for detection via FITC-labeled monoclonal antibodies that attach to the pathogen's cell wall.

Slides were examined with fluorescent microscopy for the characteristic shape and size of the water pathogens as well as the green color with positive and negative controls used as reference.

dPCR was used to detect the presence of Leptospira in the samples. dPCR protocols for Giardia and Cryptosporidium are under development



Figure 4. Giardia size is typically 10-15 um an Cryptosporidium size is typically 5 µm

## · Giardia was detected at Sketchy Bridge, a medium disturbance site as well as Herb's Garden, a high disturbance site White River Watershed sample sites did not have any positive results for Cryptosporidium and/or Giardia. No sites were positive for Leptospira via dPCR.

RESULTS

Merifluor® assays indicated that three sites were positive for Cryptosporidium and/or Giardia at Marys

Giardia and Cryptosporidium were detected at Marys River North Fork 1, a low disturbance site.

Distribution of Giardia. Cryptosporidium, and Leptospirg in the Marys and White Rivers



Figure 5. Maps of collection sites along the A. Marys River and B. White River watersheds. Color of dots medium red = bink. Black stars indicate the presence of *Giardia* and/or *Cruatesporidium* es level of disturbance

# Water Quality in the Marys and White Rivers Across a Land Use Gradient

There was no significant difference in TDS among disturbance sites (F<sub>2.35</sub>=1.985, p=0.1525), watersheds (F135=0.685, p=0.413), and the interaction between disturbance sites and watersheds (F<sub>2,35</sub>=1.407,p=0.258) (2-way ANOVA) (Figure 6).



Figure 8.

Relationship between Water Quality and Protozoal Pathogens

River Watershed (Figure 5)



Figure 8. Number of A). TDS and B). E. coli colonies measured at three sites that also tested positive for protozoal pathogens.

B.



Since few sites tested positive for protozoal pathogens, statistical

analysis is challenging, but values at each site are shown in

There was no difference between either TDS (T-test: p=0.7979)

or E. coli counts (T-test: p=0.8146) when compared to sites

positive or negative for protozoal pathogens (Figure 9).

Figure 9. Comparison of colony counts (blue) and TDS at sites either positive or negative for protozo pathogens. Error bars indicate standard deviation.

## DISCUSSION

- Waterborne pathogens are present in Marvs River with three sites testing positive.
- When PCR protocols for Giardia and Cryptosporidium are developed, we will have an additional confirmatory test so it is likely that we could find more positive sites. PCR will also enable us to determine what lineages or species of pathogens are present in the watersheds and thus allow the inference of hosts present and if humans are at risk.
- It was hypothesized that waterborne pathogens transmitted through fecal contamination would be positively correlated with E. coli counts, as a measure of fecal contamination. However, we did not find this pattern in our data, possibly due to the low number of positive sample sites.
- Marys River has higher levels of fecal contamination in the medium and low disturbance sites as compared to White River. Both rivers are similar at high disturbance sites.
- Previous year's data shows higher colony counts in 2022 than in 2023 for both watersheds but the overall pattern was similar.
- Water collection in both 2022 and 2023 was limited to 1 L per sample site. In future collections, backpack filters will be used on-site to increase sample size.
- There were only four medium and four low disturbance sites sampled at White River as compared to eleven medium and four low disturbance sites sampled at Marys River. In future collections, an equal number of sites sampled from each disturbance level will help increase the study analysis power.

# CONCLUSION

- No correlation was found between land use gradient and water pathogens Giardia and Cryptosporidium using Merifluor® immunofluorescent assay for detection.
- Leptospira was not detected along any site using dPCR so Leptospira was not used in any further data analysis.
- There is a positive relationship but no statistically significant correlation between E. coli colony counts and Giardia and Cryptosporidium in the positive sites.
- TDS values were overall greater in the negative sites as compared to the positive sites with no statistically significant correlation found.

### ACKNOWLEDGEMENTS

- Land owners who allowed access to their land/water along Marys River and White River: Starker Forests, Crestmont Land Trust, Corvallis Parks & Recreation Dept, Herb (no last name given), Dave
- Newman & company, and Harris Bridge Winery Kate Willis, Dan van Lehman, Justesen Ranch, Elizabeth Unti, Delayne Delco, Zach Harvey, Hillary and Joe
- Jensen, Mike and Laila Davis, and Mary Reechler Funding provided by WesternU Intramural Team Grant to Drs Steinauer and Hanselmann as well as OSU CCVM Biomedical Sciences.
- 2023 WesternU Student Research ONE Health Fellowship Grant funding

#### REFERENCES

Centers for Disease Control and Prevention. (2019). CDC - DPDx - Giardiasis. CDC.

https://www.cdc.gov/dpdx/giardiasis/index.html

- Humagain, S. (2021, May 4). Cryptosporidium parvum (Morphology, Life cycle, Pathogenesis, Clinical manifestation, Lab diagnosis, Treatment and Prevention and Control). Online Science Notes.
- https://onlinesciencenotes.com/cryptosporidium-parvum-morphology-life-cycle-pathogenesis-clinical-
- manifestation-lab-diagnosis-treatment-and-prevention-and-control/
- Mohammed, Dr HARAJI & Cohen, Nozha & Hakim, Karib & Aziz, Fassouane & Belahsen, Rekia. (2011). LEPTOSPIRA: Morphology, Classification and Pathogenesis. J Bacteriology and Parasitology ISSN:2155-9597 2. 2:6. 10.4172/2155-9597.1000120.
- Parasitology Review 2017. (n.d.). Www.slideshare.net. Retrieved October 22, 2023, from https://www.slideshare.net/MicrobeswithMorgan/parasitology-review-2017

Marys Rive